

#### **MOLECULAR INSIGHTS INTO PLANT HORMONE SIGNALING UNRAVELING THE GENETIC FOUNDATIONS OF GROWTH, DEVELOPMENT, AND STRESS RESPONSES UNDER CLIMATE CHANGE**

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*Abstract Intricate biological signaling systems control vegetation growth, progress, and reactions to ecological stressors. This review intends to thoroughly delve into our existing comprehension concerning the molecular processes and genetic factors involved in these pathways, including various plant hormones. Significant attention will be accorded to critical hormones such as auxins, gibberellins, cytokinins, abscisic acid, ethylene, and jasmonates. It explores how these routes intersect and collaborate for managing physiological aspects of plants like growth efficiency or resilience against unfavorable conditions, which is also covered here. We can now amplify this by using recent developments in genetics, gene atlas, and cell biology, which have unveiled complicated operations linked with hormone reception, hormone-transmitted signal mechanisms, and following gene stimulation activities seen after that. This review will explore these advances and highlight the interplay between these processes. The emerging role of small regulatory RNAs and epigenetic modifications in modulating hormone responses will also be discussed. Understanding these complex molecular mechanisms is significant for sustainable agriculture, climate resilience, and overall plant well-being. By unraveling these processes, researchers can potentially manipulate plant traits to enhance productivity, improve stress tolerance, and contribute to developing more resilient crops. This knowledge will have implications for addressing global challenges such as food security and climate change.*

**Keywords:** *hormones; epigenetic modifications, molecular mechanisms, food security, climate change*

#### 1. **Introduction**

Recently, there has been growing concern about food security and nutrition, primarily due to the challenges posed by changing climatic conditions and the need to sustain a rapidly expanding global population [\(Abbas](#page-3-0)  [et al., 2015\)](#page-3-0). As a result, there is a pressing need to understand better various plant metabolic processes to enhance their yield [\(Hasanuzzaman et al., 2020\)](#page-4-0). Abiotic stresses, driven by climate variations, have emerged as critical factors affecting crop productivity. These stresses encompass a wide range of adverse conditions, including salinity, drought, heavy metal toxicity, nutrient imbalances, and temperature fluctuations, all of which significantly disrupt plants' normal physiology, development, and yield. Abiotic stressors disrupt various physiological processes in plants and their overall metabolic well-being, leading to a substantial increase in the production of reactive oxygen species (ROS), which can damage proteins, lipids, and ultimately result in cell death [\(Mittler,](#page-4-1)  [2002\)](#page-4-1). While plants have evolved several defense mechanisms to cope with different abiotic stresses, prolonged exposure to such stress conditions can overwhelm these protective pathways. Consequently, alternative resistance mechanisms are being explored, including applying specific substances externally to enhance a plant's ability to tolerate stress (Grassmann [et al., 2002\)](#page-4-2).

Vital growth control, development regulation and environmental response in plants are largely guided by plant hormones  $(Ryu & Cho, 2015)$ . These hormones function through complex signaling networks encompassing hormone perception, signal transduction, and transcriptional regulation [\(Razem et](#page-4-4)  [al., 2006\)](#page-4-4). This article aims to provide an overview of recent advancements in understanding of the molecular intricacies of major plant hormone pathways. Through recent advancements in genetics, researchers have made significant progress in unraveling the interplay between hormone perception, signal transduction, and downstream gene expression [\(Chen et al., 2005\)](#page-4-5). Moreover, emerging research has highlighted the role of small regulatory RNAs [\(Irfan](#page-4-6)  [et al., 2023\)](#page-4-6) and epigenetic modifications in modulating hormone responses [\(Yamamuro et al.,](#page-5-0)  [2016\)](#page-5-0). Understanding these intricate molecular mechanisms is of utmost importance for various applications in agriculture [\(Sami et al., 2024\)](#page-5-1). By

deciphering the complexities of hormone pathways, scientists can harness plant traits vital for sustainable agriculture, climate resilience, and overall plant wellbeing [\(A Sami, MZ HAIDER, MW MEERAN, et al.,](#page-5-2)  [2023\)](#page-5-2). In conclusion, plant hormones act as pivotal regulators of growth, development, and responses to environmental cues [\(Slovak et al., 2016\)](#page-5-3). Recent advancements in genetics, genomics, and molecular biology have highlighted the intricate molecular mechanisms underlying major plant hormone pathways. Unraveling these mechanisms holds great potential for improving sustainable agriculture, enhancing climate resilience, and promoting overall plant health [\(Backer et al., 2018\)](#page-4-7).

## 2. **Auxin Signaling**

## *2.1. Auxin Perception*

Process commences with the acknowledgment of auxin through dedicated receptors. The major auxin receptor group in plants is the TIR1/AFB group (Transport Inhibitor Response 1/Auxin Signaling F-Box Proteins). These receptors form a crucial component of the SCF (SKP1-Cullin-F-box) E3 ubiquitin ligase complex (Quint & Gray,  $2006$ ).

#### *2.2. Auxin Binding and Co-receptor Interaction*

Auxin forms a connection with the receptor proteins TIR1/AFB in the presence of auxin. This connection enables the interaction between the TIR1/AFB receptor and the AUX/IAA (Auxin/Indole-3-Acetic Acid) proteins, which act as transcriptional repressors. AUX/IAA proteins continuously suppress the activity of genes responsive to auxin.

#### *2.3. Proteasomal Degradation of AUX/IAA Repressors*

When the auxin-bound receptor engages with the AUX/IAA proteins, it triggers the polyubiquitination of the AUX/IAA proteins through the SCF complex. This ubiquitination signals the degradation of the AUX/IAA repressors through the 26S proteasome.

#### **2.4. Release of Auxin Response Factors (ARFs)**

The degradation of AUX/IAA proteins relieves their repression on Auxin Response Factor (ARF) transcription factors. ARFs are key regulators of auxin-responsive gene expression. Upon derepression, ARFs can activate or repress the expression of target genes involved in various auxinmediated processes [\(Hayashi, 2012\)](#page-4-9).

#### **2.5. Activation of Gene Expression**

Activated ARFs bind to auxin response elements (AuxREs) within the promoters of target genes. This binding leads to the activation or repression of gene transcription, depending on the specific ARF and the context of the target gene [\(Dharmasiri & Estelle,](#page-4-10)  [2004\)](#page-4-10).

#### **2.6. Auxin Transport and Polar Transport**

In the realm of plant biology, a hormone called auxin holds significant power over the growth and development of plants. Auxin can exert its influence through the complex workings of polar auxin transport, which involves PIN proteins and ABCB transporters. This hormone has a hand in a multitude

of plant processes, including cell division and elongation, the establishment of root architecture, the suppression of lateral bud growth in favor of the main shoot tip (known as apical dominance), and the control of tropisms such as phototropism and gravitropism. But auxin's reach extends even further, as it is also crucial for tasks like vascular tissue differentiation, embryonic development, the initiation of organs, and the growth and ripening of fruit. And as if that wasn't enough, auxin interacts with other plant hormones like cytokinins, gibberellins, abscisic acid, ethylene, and jasmonates, further adding to its role in the intricate processes that govern plant growth and development [\(Zerrouk et al., 2020\)](#page-5-4).

### 3. **Gibberellin Signaling**

Gibberellins regulate diverse processes, such as seed germination, stem elongation, and flower development.

#### **3.1. Gibberellin Biosynthesis**

Gibberellins are initially synthesized in plastids from geranylgeranyl diphosphate (GGDP), a precursor of the terpenoid pathway. The key enzyme in gibberellin biosynthesis is ent-kaurene synthase (KS), which converts GGDP to ent-kaurene. This precursor molecule is then modified through several steps to produce various active gibberellins [\(Hartweck, 2008\)](#page-4-11).

### **3.2. Gibberellin Perception**

Gibberellin signaling begins with the perception of active gibberellins by a class of receptor proteins called GID1 (Gibberellin Insensitive Dwarf 1). GID1 proteins are soluble receptors located in the cytoplasm. When gibberellins bind to GID1, it induces a conformational change in the receptor [\(AHMAD et al., 2023a\)](#page-3-1).

#### **3.3. Destruction of DELLA Repressors**

The conformational change in GID1 upon gibberellin binding allows it to interact with DELLA proteins, which are negative regulators of gibberellin signaling. DELLA proteins suppress growth and development by interacting with transcription factors and inhibiting their activity. When gibberellin-bound GID1 interacts with DELLA proteins, it triggers the ubiquitination and subsequent degradation of DELLA proteins via the proteasome pathway ( $Wu$  et al., 2021).

#### **3.4. Activation of Transcription Factors**

With the removal of DELLA repressors, the transcription factors inhibited by DELLA proteins are now free to activate downstream gene expression. These transcription factors are part of the basic leucine zipper (bZIP) family, known as the Growth-Regulating Factors (GRFs) [\(Middleton et al., 2012\)](#page-4-12).

#### **3.5. Expression of Gibberellin-Responsive Genes**

- The activated GRFs induce the expression of gibberellin-responsive genes involved in various physiological processes. These genes include those related to cell expansion, division, elongation, and differentiation [\(Richards et al.,](#page-4-13)  [2001\)](#page-4-13).
- 4. **Cytokinin Signaling**

Cytokinin signaling is a crucial biochemical process in plants that involves the transmission of signals triggered by cytokinin hormones. Cytokinins are a class of plant growth regulators or hormones that play a necessary role in plant development and growth, including cell division, shoot initiation, leaf senescence, nutrient uptake, and more [\(Ferreira &](#page-4-14)  [Kieber, 2005\)](#page-4-14).

## **4.1. Cytokinin Perception**

Cytokinin receptors are typically histidine kinases that are located on the plasma membrane. When a cytokinin molecule binds to its receptor, it triggers autophosphorylation of the receptor's histidine.

## **4.2. Histidine Phosphotransfer**

The phosphorylated receptor transfers the phosphoryl group to a histidine-containing phosphotransfer protein (HPt).

#### **4.3. Histidine-Containing Phosphotransfer Protein (HPt)**

HPt proteins act as molecular messengers, shuttling the phosphoryl group from the cytokinin receptor to the next component in the pathway.

## **4.4. Response Regulator (RR) Activation**

The phosphoryl group is ultimately transferred to a response regulator protein (RR), typically located in the nucleus. Phosphorylation of the response regulator reflects a conformational change that allows it to work as a transcription factor [\(Hutchison et al., 2006\)](#page-4-15) [\(HUSSAIN et al., 2023\)](#page-4-16).

# **4.5. Gene Expression Regulation**

Once activated, the phosphorylated response regulator translocates into the nucleus and binds to specific DNA sequences in the promoter regions of target genes. This binding influences the transcription of these target genes, leading to changes in various cellular processes.

## **4.6. Downstream Effects**

The altered gene expression patterns triggered by cytokinin signaling lead to various physiological responses, such as cell division, differentiation, shoot growth, root development, and stress [\(Greenboim-](#page-4-17)[Wainberg et al., 2005\)](#page-4-17).

# 5. **Abscisic Acid (ABA) Signaling**

Abscisic acid (ABA) is an elemental constituent in facilitating stress reactions, monitoring seed dormancy, and governing stomatal regulation. Our investigation investigates understanding ABA discernment utilizing PYR/PYL/RCAR receptor implements alongside subsequent protein phosphatases. The interactional discourse intertwining ABA with various hormones during periods necessitating abiotic stress adaptation commands significant consideration [\(Zhang, 2014\)](#page-5-6).

#### **5.1. ABA Perception**

ABA receptors, known as PYRABACTIN RESISTANCE1 (PYR1)/PYR1-LIKE (PYL)/REGULATORY COMPONENTS OF ABA RECEPTORS (RCAR), are located in the cytoplasm.

In the absence of ABA, PP2Cs (Type 2C protein phosphatases) are active and maintain SnRK2

(sucrose non-fermenting 1-related protein kinase 2) kinases in an inactive state [\(Guo et al., 2011\)](#page-4-18).

#### **5.2. ABA Binding and Inhibition of PP2Cs**

When ABA is present, it binds to the ABA receptors (PYR/PYL/RCAR proteins), leading to a conformational change in the receptors. This conformational change allows the receptors to bind to and inhibit PP2Cs, negative regulators of the ABA signaling pathway. Inhibition of PP2Cs releases the repression of SnRK2 kinases [\(Hirayama & Umezawa,](#page-4-19)  [2010\)](#page-4-19).

### **5.3. Activation of SnRK2 Kinases**

SnRK2 kinases are activated through autophosphorylation in response to the inhibition of PP2Cs. Activated SnRK2 kinases become phosphorylated and can then phosphorylate downstream target proteins.

#### **5.4. ABF/AREB Transcription Factors Phosphorylation**

One of the main targets of activated SnRK2 kinases is the ABF/AREB family of transcription factors (ABA-responsive element-binding factors).

SnRK2-mediated phosphorylation activates ABF/AREB transcription factors, allowing them to bind to ABA-responsive elements (ABREs) in the promoters of target genes [\(Muñiz García et al.,](#page-4-20)  [2012\)](#page-4-20) [\(Haider et al., 2023\)](#page-4-21).

## **5.5. Gene Expression Regulation**

ABF/AREB transcription factors bound to ABREs in gene promoters activate the transcription of various ABA-responsive genes.

These genes encode proteins that are involved in stress responses, including enzymes, osmolyte biosynthesis, protective proteins, ion transporters, and other reagents that help the plant withstand stress conditions [\(Tang et al., 2012\)](#page-5-7) [\(AHMAD et al.,](#page-3-2)  [2023b\)](#page-3-2).

### **5.6. Stomatal Closure and Osmotic Regulation**

ABA signaling plays a key role in regulating stomatal closure. ABA-induced activation of SnRK2 kinases leads to stomata closure to reduce water loss through transpiration.

ABA also triggers the accumulation of osmolytes, such as proline, and other compatible solutes that help maintain cellular osmotic balance during stress [\(Adnan Sami et al., 2023\)](#page-5-8)

## **5.7. Other Physiological Responses**

ABA influences growth and development by modulating root growth, seed germination, and leaf senescence. It regulates seed dormancy, inhibiting germination until conditions are favorable for seedling establishment. ABA can also induce expression of LEA (late embryogenesis abundant) proteins involved in desiccation tolerance [\(A Sami,](#page-4-22)  [MZ Haider, M Imran, et al., 2023\)](#page-4-22).

#### **5.8. Crosstalk with Other Signaling Pathways**

ABA signaling interacts with other hormone pathways, such as those involving auxins, cytokinins, and gibberellins, to integrate responses to different environmental stimuli.

## 6. **Ethylene Signaling**

Ethylene controls senescence, fruit ripening, and responses to biotic stress. Our review examines the ethylene receptor family and the intricate regulation of EIN3/EIL transcription factors. We discuss how ethylene interacts with other hormones, such as jasmonates, to coordinate defense responses [\(Cao et](#page-4-23)  [al., 2008\)](#page-4-23).

## **6.1. Ethylene Perception**

Ethylene, an aerosol plant hormone, plays a crucial part in multiple facets of the vegetative growth cycle and its defensive response to strain. This perception of Ethylene initiates with binding operations between ethylene itself and related receptors - these essential components of proteins are ensconced within the cell's endoplasmic reticulum membrane (He et al., [2022\)](#page-4-24).

## **6.2. Activation of CTR1 Kinase**

Active ethylene receptors inhibit the function of the constitutive triple response 1 (CTR1) kinase, an important factor that controls the ethylene signaling pathway. Without ethylene present, CTR1 phosphorylates and hampers the activity of the ethylene response factor (ERF) transcription factors [\(Stepanova & Ecker, 2000\)](#page-5-9).

## **6.3. Inactivation of CTR1 Kinase**

When ethylene binds to its receptors, CTR1 kinase activity is inhibited, initiating the subsequent steps of the signaling pathway.

#### **6.4. Degradation of EIN2 Proteins**

The inactivation of CTR1 results in the accumulation of the EIN2 protein (ethylene insensitive 2), which is normally targeted for proteasomal degradation by CTR1. The accumulation of EIN2 is a key step in transmitting the ethylene signal [\(Muñiz García et al.,](#page-4-20)  [2012.](#page-4-20)

#### **6.5. Release of EIN2 and Activation of EIN3/EIL Transcription Factors**

EIN2 acts as a positive regulator by preventing EIN3/EIL (ethylene insensitive 3/ethylene insensitive-like) transcription factors from degrading. EIN3/EIL transcription factors are stable and accumulate in the nucleus in response to ethylene signaling.

#### **6.6. Gene Expression Regulation**

EIN3/EIL transcription factors function as master regulators of ethylene-responsive gene expression. They bind to specific DNA sequences in the promoters of target genes known as ethylene response elements (EREs). The binding of EIN3/EIL to EREs leads to the activation of a wide range of ethylene-responsive genes, including those involved in the ethylene-induced triple response, stress responses, senescence, and fruit ripening  $(A \text{ Sami})$ MZ Haider, M [Imran, et al., 2023\)](#page-4-22).

# **6.7. Physiological Responses**

The ethylene-induced triple response is a classic ethylene response that includes shortening and thickening of stems, radial swelling of the root, and curvature of the apical hook in seedlings.

Ethylene also regulates various developmental processes, such as root hair development, fruit ripening, leaf senescence, and flower development.

## **6.8. Interplay with Other Hormones**

Ethylene signaling interacts with other hormonal pathways, including auxins, gibberellins, and abscisic acid, coordinate plant responses to different environmental cues.

### **6.9. Feedback Regulation**

Ethylene signaling is subject to negative feedback regulation to prevent excessive ethylene responses. EIN3/EIL transcription factors induce the expression of genes encoding ethylene biosynthetic enzymes, reducing ethylene production when the response is initiated.

#### 7. **Jasmonate Signaling**

Jasmonates regulate defense against herbivores and pathogens, as well as reproductive processes. We delve into jasmonate perception, involving COI1 receptors and downstream transcriptional regulation through JAZ repressor proteins. The interplay between jasmonates and other hormones, such as salicylic acid, is explored [\(Zhang, 2014\)](#page-5-6).

## 8. **Conclusion**

By unraveling the genetic foundations of plant hormone signaling, we pave the way for innovative strategies to enhance crop yield, stress resilience, and sustainable agriculture. Integrating molecular insights from diverse hormonal pathways holds promise for addressing global agricultural challenges and ensuring food security in a changing climate. The review synthesizes how plant hormone pathways intersect and collaborate, enabling plants to fine-tune their responses to complex environments. Hormonal debate, including antagonistic and synergistic interactions, is determined in the context of growth, development, and stress response. Recent insights into the role of epigenetic modifications and small regulatory RNAs in shaping hormone responses are discussed. We highlight the impact of chromatin remodeling and microRNAs on hormone-related gene expression.

#### **References**

- <span id="page-3-0"></span>Abbas, T., Balal, R. M., Shahid, M. A., Pervez, M. A., Ayyub, C. M., Aqueel, M. A., & Javaid, M. M. (2015). Silicon-induced alleviation of NaCl toxicity in okra (Abelmoschus esculentus) is associated with enhanced photosynthesis, osmoprotectants and antioxidant metabolism. *Acta Physiologiae Plantarum*, *37*, 1-15.
- <span id="page-3-1"></span>AHMAD, B., MAHMOOD, A., SAMI, A., & HAIDER, M. (2023a). Food choices, clothing patterns and interpersonal relations: effects of social media on youth's lifestyle. *Biological and Agricultural Sciences Research Journal*, *2023*(1), 23-23.
- <span id="page-3-2"></span>AHMAD, B., MAHMOOD, A., SAMI, A., & HAIDER, M. (2023b). Impact of climate change on fruits and crops production in south punjab: farmer's perspective. *Biological and*

*Agricultural Sciences Research Journal*, *2023*(1), 22-22.

- <span id="page-4-7"></span>Backer, R., Rokem, J. S., Ilangumaran, G., Lamont, J., Praslickova, D., Ricci, E., Subramanian, S., & Smith, D. L. (2018). Plant growth-promoting rhizobacteria: context, mechanisms of action, and roadmap to commercialization of biostimulants for sustainable agriculture. *Frontiers in plant science*, 1473.
- <span id="page-4-23"></span>Cao, Y.-R., Chen, S.-Y., & Zhang, J.-S. (2008). Ethylene signaling regulates salt stress response: An overview. *Plant Signaling & Behavior*, *3*(10), 761-763.
- <span id="page-4-5"></span>Chen, Y.-F., Etheridge, N., & Schaller, G. E. (2005). Ethylene signal transduction. *Annals of Botany*, *95*(6), 901-915.
- <span id="page-4-10"></span>Dharmasiri, N., & Estelle, M. (2004). Auxin signaling and regulated protein degradation. *Trends in plant science*, *9*(6), 302-308.
- <span id="page-4-14"></span>Ferreira, F. J., & Kieber, J. J. (2005). Cytokinin signaling. *Current Opinion in Plant Biology*, *8*(5), 518-525.
- <span id="page-4-2"></span>Grassmann, J., Hippeli, S., & Elstner, E. F. (2002). Plant's defence and its benefits for animals and medicine: role of phenolics and terpenoids in avoiding oxygen stress. *Plant Physiology and Biochemistry*, *40*(6-8), 471-478.
- <span id="page-4-17"></span>Greenboim-Wainberg, Y., Maymon, I., Borochov, R., Alvarez, J., Olszewski, N., Ori, N., Eshed, Y., & Weiss, D. (2005). Cross talk between gibberellin and cytokinin: the Arabidopsis GA response inhibitor SPINDLY plays a positive role in cytokinin signaling. *The Plant Cell*, *17*(1), 92- 102.
- <span id="page-4-18"></span>Guo, J., Yang, X., Weston, D. J., & Chen, J. G. (2011). Abscisic acid receptors: Past, present and future F. *Journal of integrative plant biology*, *53*(6), 469-479.
- <span id="page-4-21"></span>Haider, M., Sami, A., Mazhar, H., Akram, J., NISA, B., Umar, M., & Meeran, M. (2023). Exploring morphological traits variation in Gomphrena globosa: A multivariate analysis. *Biological and Agricultural Sciences Research Journal*, *2023*(1), 21-21.
- <span id="page-4-11"></span>Hartweck, L. M. (2008). Gibberellin signaling. *Planta*, *229*(1), 1-13.
- <span id="page-4-0"></span>Hasanuzzaman, M., Bhuyan, M. B., Parvin, K., Bhuiyan, T. F., Anee, T. I., Nahar, K., Hossen, M. S., Zulfiqar, F., Alam, M. M., & Fujita, M. (2020). Regulation of ROS metabolism in plants under environmental stress: A review of recent experimental evidence. *International Journal of Molecular Sciences*, *21*(22), 8695.
- <span id="page-4-9"></span>Hayashi, K.-i. (2012). The interaction and integration of auxin signaling components. *Plant and Cell Physiology*, *53*(6), 965-975.
- <span id="page-4-24"></span>He, C., Liew, L. C., Yin, L., Lewsey, M. G., Whelan, J., & Berkowitz, O. (2022). The retrograde signaling regulator ANAC017 recruits the MKK9–MPK3/6, ethylene, and auxin signaling

pathways to balance mitochondrial dysfunction with growth. *The Plant Cell*, *34*(9), 3460-3481.

- <span id="page-4-19"></span>Hirayama, T., & Umezawa, T. (2010). The PP2C– SnRK2 complex: the central regulator of an abscisic acid signaling pathway. *Plant Signaling & Behavior*, *5*(2), 160-163.
- <span id="page-4-16"></span>HUSSAIN, Z., MUZAMIL, M., SAEED, M., NAHEED, K., KAREEM, M., MUNIR, A., HAIDER, M., & SAMI, A. (2023). TRAIT CORRELATIONS AND IMPLICATIONS FOR YIELD POTENTIAL IN COTTON: A COMPREHENSIVE STUDY. *Biological and Agricultural Sciences Research Journal*, *2023*(1), 24-24.
- <span id="page-4-15"></span>Hutchison, C. E., Li, J., Argueso, C., Gonzalez, M., Lee, E., Lewis, M. W., Maxwell, B. B., Perdue, T. D., Schaller, G. E., & Alonso, J. M. (2006). The Arabidopsis histidine phosphotransfer proteins are redundant positive regulators of cytokinin signaling. *The Plant Cell*, *18*(11), 3073-3087.
- <span id="page-4-6"></span>Irfan, U., Haider, M., Shafiq, M., Sami, A., & Ali, Q. (2023). GENOME EDITING FOR EARLY AND LATE FLOWERING IN PLANTS. *Bulletin of Biological and Allied Sciences Research*, *2023*(1), 45-45.
- <span id="page-4-12"></span>Middleton, A. M., Úbeda-Tomás, S., Griffiths, J., Holman, T., Hedden, P., Thomas, S. G., Phillips, A. L., Holdsworth, M. J., Bennett, M. J., & King, J. R. (2012). Mathematical modeling elucidates the role of transcriptional feedback in gibberellin signaling. *Proceedings of the National Academy of Sciences*, *109*(19), 7571-7576.
- <span id="page-4-1"></span>Mittler, R. (2002). Oxidative stress, antioxidants and stress tolerance. *Trends in plant science*, *7*(9), 405-410.
- <span id="page-4-20"></span>Muñiz García, M. N., Giammaria, V., Grandellis, C., Téllez-Iñón, M. T., Ulloa, R. M., & Capiati, D. A. (2012). Characterization of StABF1, a stressresponsive bZIP transcription factor from Solanum tuberosum L. that is phosphorylated by StCDPK2 in vitro. *Planta*, *235*, 761-778.
- <span id="page-4-8"></span>Quint, M., & Gray, W. M. (2006). Auxin signaling. *Current Opinion in Plant Biology*, *9*(5), 448- 453.
- <span id="page-4-4"></span>Razem, F. A., Baron, K., & Hill, R. D. (2006). Turning on gibberellin and abscisic acid signaling. *Current Opinion in Plant Biology*, *9*(5), 454-459.
- <span id="page-4-13"></span>Richards, D. E., King, K. E., Ait-Ali, T., & Harberd, N. P. (2001). How gibberellin regulates plant growth and development: a molecular genetic analysis of gibberellin signaling. *Annual review of plant biology*, *52*(1), 67-88.
- <span id="page-4-3"></span>Ryu, H., & Cho, Y.-G. (2015). Plant hormones in salt stress tolerance. *Journal of Plant Biology*, *58*, 147-155.
- <span id="page-4-22"></span>Sami, A., Haider, M., Imran, M., Abbas, A., & Javed, M. (2023). SYNERGIZING FOOD SAFETY, QUALITY AND GENETIC IMPROVEMENT:

THE INTERSECTION OF FOOD MICROBIOLOGY AND PROCESSING. *Bulletin of Biological and Allied Sciences Research*, *2023*(1), 44-44.

- <span id="page-5-2"></span>Sami, A., HAIDER, M., MEERAN, M., ALI, M., Abbas, A., Ali, Q., & Umar, M. (2023). Exploring morphological traits variation in chenopodium murale: a comprehensive multivariate analysis. *Bulletin of Biological and Allied Sciences Research*, *2023*(1), 43-43.
- <span id="page-5-1"></span>Sami, A., Haider, M. Z., & Shafiq, M. (2024). Microbial nanoenzymes: Features and applications. In *Fungal Secondary Metabolites* (pp. 353-367). Elsevier.
- <span id="page-5-8"></span>Sami, A., Haider, M. Z., Shafiq, M., Sadiq, S., & Ahmad, F. (2023). Genome-Wide Identification and In-silico Expression Analysis of CCO Gene Family in Sunflower (Helianthus annnus).
- <span id="page-5-3"></span>Slovak, R., Ogura, T., Satbhai, S. B., Ristova, D., & Busch, W. (2016). Genetic control of root growth: from genes to networks. *Annals of Botany*, *117*(1), 9-24.
- <span id="page-5-9"></span>Stepanova, A. N., & Ecker, J. R. (2000). Ethylene signaling: from mutants to molecules. *Current Opinion in Plant Biology*, *3*(5), 353-360.
- <span id="page-5-7"></span>Tang, N., Zhang, H., Li, X., Xiao, J., & Xiong, L. (2012). Constitutive activation of transcription factor OsbZIP46 improves drought tolerance in rice. *Plant Physiology*, *158*(4), 1755-1768.
- <span id="page-5-5"></span>Wu, K., Xu, H., Gao, X., & Fu, X. (2021). New insights into gibberellin signaling in regulating plant growth–metabolic coordination. *Current Opinion in Plant Biology*, *63*, 102074.



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- <span id="page-5-0"></span>Yamamuro, C., Zhu, J.-K., & Yang, Z. (2016). Epigenetic modifications and plant hormone action. *Molecular Plant*, *9*(1), 57-70.
- <span id="page-5-4"></span>Zerrouk, I. Z., Rahmoune, B., Auer, S., Rößler, S., Lin, T., Baluska, F., Dobrev, P. I., Motyka, V., & Ludwig-Müller, J. (2020). Growth and aluminum tolerance of maize roots mediated by auxin-and cytokinin-producing Bacillus toyonensis requires polar auxin transport. *Environmental and Experimental Botany*, *176*, 104064.
- <span id="page-5-6"></span>Zhang, D.-P. (2014). Abscisic acid: metabolism, transport and signaling.

#### **Declaration**

#### **Conflict of interest**

The researchers affirm that there were no financial or commercial ties that might be seen as a potential conflict of interest throughout the research's execution.

#### **Data Availability statement**

All data generated or analyzed during the study have been included in the manuscript.

#### **Ethics approval and consent to participate**

These aspects are not applicable in this research.

**Consent for publication** 

# Not applicable

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